THE CITY OF SAN DIEGO

# EMTS Division Laboratory Quality Assurance Report 2010

# City of San Diego Ocean Monitoring Program

**Public Utilities Department**
**Environmental Monitoring and Technical Services Division**

# EMTS Division Laboratory Quality Assurance Report 2010

**Prepared by:**

**City of San Diego**
**Ocean Monitoring Program**
**Public Utilities Department**
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# 2010 Quality Assurance Report

# Introduction

The Quality Assurance/Quality Control (QA/QC) Program for the Environmental Monitoring and Technical Services (EMTS) Division Laboratory, Public Utilities Department, City of San Diego (City) includes various practices that have been instituted to ensure the accuracy and reliability of ocean monitoring data reported to regulatory agencies in compliance with the reporting requirements specified in several National Pollutant Discharge Elimination System (NPDES) permits (Table 1). These QA/QC procedures assure the quality of field sampling, laboratory analysis, records keeping, data entry, electronic data collection/transfer, as well as data analysis and reporting. The procedures are regularly reviewed and updated to reflect ongoing changes in NPDES permit requirements, sample collection, methods, technology, and applicability of new analytical methods.

The comprehensive QA/QC activities of the EMTS Division Laboratory are documented separately in the laboratory's Quality Assurance Project Plan, which is currently under revision (City of San Diego, in prep). Additionally, the EMTS Division maintains certification through the International Organization for Standardization (ISO) 14001 Environmental Management Systems program (ISO 14001). As part of continuation in the ISO certification process, EMTS underwent and passed an external audit in 2010 conducted by a third-party auditor. This report summarizes the QA/QC activities that were conducted during calendar year 2010 by EMTS staff in support of NPDES permit mandated monitoring for the City's Point Loma Wastewater Treatment Plant and South Bay Water Reclamation Plant, as well as similar activities required for the International Wastewater Treatment Plant operated by the International Boundary and Water Commission.

# Facilities and Staff

The EMTS Division includes three sections (laboratories) that participate in the receiving waters monitoring activities associated with the above NPDES permits: (1) Marine Biology and Ocean Operations; (2) Microbiology; (3) Wastewater Chemistry Services. The Marine Biology and Ocean Operations section (Marine Biology Lab) and Marine Microbiology and Vector Management section (Marine Microbiology Lab) are located at the EMTS Division Laboratory (2392 Kincaid Road, San Diego, CA 92101). These two laboratories are responsible for conducting most field operations and performing subsequent biological and oceanographic assessments associated with the City's Ocean Monitoring Program (e.g., water quality, benthic sediments and macrofauna, trawl-caught fishes and invertebrates, contaminant bioaccumulation in fishes). Marine Biology and Microbiology personnel are organized into technical work groups based on their major work responsibilities and areas of expertise (see Appendix A.1 and A.2). Brief descriptions of the areas of emphasis for each work group are given in the following sections.

The Wastewater Chemistry Services (WCS) section is located at other City facilities and performs chemical analyses of the various seawater, sediment and fish tissue samples collected by the program. Descriptions of the WCS organization (Appendix A.3) and additional quality assurance procedures conducted in support of the receiving waters monitoring programs are presented in a separate QA report each year (e.g., City of San Diego 2011).

**Table 1**
National Pollutant Discharge Elimination System (NPDES) permits subject to receiving waters monitoring by the EMTS Division Laboratories.

| Facility | Owner/Operator | NPDES Permit No | Effective Date | Comment |
|---|---|---|---|---|
| Point Loma Wastewater Treatment Plant | City of San Diego | CA0107409, Order No. R9-2009-0001 | August 1, 2010 | Replaced Order No. R9-2002-0025 |
| South Bay Water Reclamation Plant | City of San Diego | CA0109045, Order No. R9-2006-0067 | January 1, 2007 | |
| International Wastewater Treatment Plant | International Boundary and Water Commission | CA0108928, Order No. 96-50 | November 14, 1996 | |

## Marine Biology and Ocean Operations

***Data Management and Reporting (DM&R)***: The primary responsibility of the DM&R work group is to analyze and report receiving waters monitoring data. This includes data QA, data analysis, and the interpretation of results from the receiving waters monitoring activities and other contract work. DM&R personnel work closely with the IT/GIS group (described below) to perform QA of all receiving waters monitoring data that is entered into the laboratory's database. Various software packages for data management, data manipulation, statistical analysis, and presentation are used to manage and analyze data from every aspect of receiving waters monitoring. Interpretation of these analyses is reported to regulatory and contract agencies in the form of monthly and annual reports.

***Information Technology and Geographic Information Systems (IT/GIS)***: The IT/GIS work group is primarily responsible for the administration of the lab's database, performing geospatial data analysis, and generating all map products as needed for the ocean monitoring program. Daily responsibilities include entry and archiving of ocean monitoring data, validation of data accuracy, maintenance of database structure and integrity, oversight of database access/security issues, and management of database enhancements. This group is also responsible for IT project planning, workflow automation programming, and website maintenance to support Marine Biology and other EMTS laboratory staff.

***Ocean Operations***: This work group is comprised of two subsections, Ocean Operations and Vessel Operations. Ocean Operations personnel oversee and conduct water quality sampling, benthic sediment and macrofauna sampling, trawling and rig-fishing, diving operations, and ocean outfall inspections. These staff maintain and calibrate all oceanographic instrumentation, SCUBA equipment, and the laboratory's remotely operated vehicle (ROV). Vessel Operations personnel are primarily responsible for the operation and maintenance of the City's two monitoring vessels, the 48' *Oceanus* and the 42' *Monitor III*. When in port, the group's Boat Operators schedule and oversee all regular vessel maintenance as well as any modifications that may become necessary. While at sea, they are responsible for ensuring the safety of the crew, locating and maintaining position at the monitoring stations, and assisting with various deck activities during field operations.

***Taxonomy***: The Taxonomy work group coordinates and manages the processing of all benthic macrofauna and trawl invertebrate samples, maintains the taxonomic literature and voucher collections, and conducts taxonomic training. In addition, they produce in-house identification sheets and keys

**Table 2**
ELAP certifications for the Environmental Monitoring and Technical Services Division Laboratories.

| ELAP Laboratory | Address | Phone | ELAP Code | Cert.No. |
|---|---|---|---|---|
| Marine Microbiology | 2392 Kincaid Road<br>San Diego, CA<br>92101-0811 | 619-758-2360 | CA01393 | 2185 |
| Toxicology | 2392 Kincaid Road<br>San Diego, CA<br>92101-0811 | 619-758-2348 | CA01302 | 1989 |

to important species and other taxa. Members of this group participate in a regional taxonomic standardization program and perform all QA/QC procedures to ensure the accuracy of the taxonomic identifications made by laboratory personnel.

***Toxicology***: The Toxicology Laboratory is certified by the State of California Department of Health Services, Environmental Laboratory Accreditation Program (ELAP), which is renewed on a biennial basis. The current ELAP certification is scheduled for renewal on April 30, 2012 (Table 2). Toxicology personnel are responsible for conducting all acute and chronic toxicity testing required by the City's NPDES permits and contractual obligations. Primary responsibilities include collection of wastewater effluent or other types of samples, maintaining test organisms and laboratory supplies, calibration of test instruments, conducting acute and chronic bioassays, record keeping, and the statistical evaluation, interpretation and reporting of all toxicology data. In addition, the Toxicology Lab maintains a separate Quality Assurance Manual in accordance with ELAP requirements, which contains up-to-date revisions to reflect current laboratory practices and procedures, and to ensure timely document version control.

## Marine Microbiology and Vector Management

***Marine Microbiology***: The Marine Microbiology Laboratory (MML) is also certified by ELAP, with the current certification in effect until November 30, 2012 (Table 2). The MML is responsible for the quantification and identification of bacteria found in environmental samples. Responsibilities include the preparation of microbiological media, reagents, sample bottles, supplies and equipment, the collection of field samples along the shore, and a variety of analyses (e.g., membrane filtration, multiple tube fermentation, and Colilert-18 and Enterolert chromogenic substrate analyses) as appropriate to the sample type and as required by the NPDES permits. In addition, the group is responsible for the physical maintenance and quality assurance of large instruments such as autoclaves, incubators, water baths, ultra-freezers, bacteriological safety cabinet, and three reagent-grade water point-of-use systems. Members are also responsible for developing sampling, analytical, and quality assurance protocols for special projects or studies involving microbiology.

***Vector Management***: The Vector Management group provides for monitoring, surveillance, control and prevention of insects and other pests that are capable of transmitting diseases or causing harm to humans. The primary methods of control include environmental conservation measures, education, and water management techniques aided by appropriate chemical and biological control technology. The vector control program uses methods to census animal populations to determine control effectiveness and trends. Areas of responsibility include treatment plants, pump stations, buildings and office facilities. Biological assessments (bioassessments) of urban creeks and streams are conducted to evaluate and analyze short and long-term impacts of sewage spills into watersheds and receiving waters.

Field samples of aquatic communities are collected and field water quality indicators are measured. Physical habitat characteristics and anthropogenic changes are evaluated. Measures, evaluations, and comparisons are made to yield relative ratings of conditions within a specified community.

## Scope of Work

Treated effluent from the Point Loma Wastewater Treatment Plant (PLWTP) is discharged to the Pacific Ocean through the Point Loma Ocean Outfall (PLOO), whereas the South Bay Ocean Outfall (SBOO) accepts treated effluent from the South Bay Water Reclamation Plant (SBWRP) and International Wastewater Treatment Plant (IWTP). The separate NPDES permits associated with each of these treatment facilities define the requirements for toxicity testing and the monitoring of receiving waters for each discharge site. The permits define the sampling plans, compliance criteria, laboratory analyses, statistical analyses and reporting guidelines.

The core receiving waters monitoring efforts for both the Point Loma and South Bay monitoring programs are summarized in Tables 3 and 4, while the fixed-grid sampling sites for each program are shown in Figure 1. These core monitoring activities include weekly sampling of seawater from recreational areas along the shoreline and within the Point Loma and Imperial Beach kelp beds, as well as monthly or quarterly offshore sampling in order to document water quality conditions in the region. Benthic samples are collected semiannually or annually to monitor sediment conditions and macrofaunal communities. Trawl surveys are performed quarterly in the South Bay region and semiannually off Point Loma to monitor the ecological health of demersal fish and epibenthic invertebrate communities. Additionally, fish tissue samples are collected and analyzed on either a semiannual or annual basis to monitor levels of chemical constituents that may have ecological or human health implications. Toxicity testing consists of acute and chronic bioassays of influent, effluent, and groundwater samples. The general toxicity testing required by the NPDES permits is outlined in Table 5. The results of these receiving waters monitoring activities and toxicity tests are analyzed and presented in various regulatory reports that are submitted to the San Diego Regional Water Quality Control Board (RWQCB).

In addition to the above core monitoring efforts, the City also conducts "strategic process studies" (i.e., special projects) as part of its regulatory requirements and as defined by the Model Monitoring Program developed for large ocean dischargers in southern California (Schiff et al. 2001). These special studies are determined by the City in coordination with the RWQCB and the United States Environmental Protection Agency (U.S. EPA), and are generally designed to address recommendations for enhanced environmental monitoring of the San Diego coastal region put forth in a peer-reviewed report coordinated by scientists at the Scripps Institution of Oceanography (SIO 2004). Data for these directed studies are subject to similar QA/QC procedures as the routine monitoring data, although the projects themselves do not necessarily conform to the same analysis and reporting schedules. Thus, details and results of ongoing QA/QC activities associated with these special studies are not included in this report unless otherwise indicated.

As part of its regulatory requirements, the City also participates in regional monitoring activities for the entire Southern California Bight coordinated by the Southern California Coastal Water Research Project (SCCWRP). The intent of the regional programs is to maximize the efforts of the various partner agencies (e.g., municipal dischargers, research agencies) using a more cost-effective monitoring design and to best utilize the pooled scientific resources of the region. These bight-wide surveys have included the 1994 Southern California Bight Pilot Project (SCBPP) and subsequent

# Table 3

NPDES-permit mandated receiving waters sampling effort for the Point Loma Ocean Outfall Monitoring Program, excluding resamples, QA/QC analyses (e.g., duplicate/split samples), or special studies.

| Monitoring Component | Location | Number of Stations/Zones | Sample Type | Discrete No. Samples/Site | Sampling Frequency | Sampling Times/Yr | Discrete No. Samples/Yr | Parameters | No. "Samples" Analyzed/Yr | Notes |
|---|---|---|---|---|---|---|---|---|---|---|
| Water Quality | shore | 8 | Seawater - Bacti | 1 | weekly | 52 | 416 | T, F, E [a] | 1248 | 1 sample/station |
| *Microbiology* | kelp | 8 | Seawater - Bacti | 3 | 5x/month | 60 | 1440 | T, F, E [a] | 4320 | 3 depths/station |
| & | | 8 | Seawater - $NH_4$ | 3 | quarterly | 4 | 96 | $NH_4$ | 96 | 3 depths/station/quarter |
| *Oceanographic Conditions* | | 8 | CTD | 1 | 5x/month | 60 | 480 | CTD profile [c] | 3840 | 1 cast/station |
| | offshore | 3 | Seawater - Bacti | 3 | quarterly | 4 | 36 | E [b] | 36 | 3 depths/station (18m stns) |
| | (n=36) | 11 | Seawater - Bacti | 3 | quarterly | 4 | 132 | E [b] | 132 | 3 depths/station (60m stns) |
| | | 11 | Seawater - Bacti | 4 | quarterly | 4 | 176 | E [b] | 176 | 4 depths/station (80m stns) |
| | | 11 | Seawater - Bacti | 5 | quarterly | 4 | 220 | E [b] | 220 | 5 depths/station (98m stns) |
| | | 3 | Seawater - $NH_4$ | 3 | quarterly | 4 | 36 | $NH_4$ | 36 | 3 depths/stn (18m stns, State Waters) |
| | | 9 | Seawater - $NH_4$ | 3 | quarterly | 4 | 108 | $NH_4$ | 108 | 3 depths/stn (60m stns, State Waters) |
| | | 3 | Seawater - $NH_4$ | 4 | quarterly | 4 | 48 | $NH_4$ | 48 | 4 depths/stn (80m stns, State Waters) |
| | | 36 | CTD | 1 | quarterly | 4 | 144 | CTD profile [d] | 1296 | 1 cast/station |
| Sediment Quality | offshore | 22 | Grab | 1 | semiannual | 2 | 44 | sediment constituents [e] | 396 | 1 grab/station (Jan, Jul) |
| Benthic Macrofauna | offshore | 22 | Grab | 2 | semiannual | 2 | 88 | community structure | 88 | 2 replicate grabs/station (Jan, Jul) |
| Demersal Fishes & Invertebrates | offshore | 6 | Trawl | 1 | semiannual | 2 | 12 | community structure | 12 | 1 trawl/station (Jan, Jul) |
| Bioaccumulation | offshore | 4 | Trawl | 3 | annual | 1 | 12 | liver tissue contaminants [f] | 48 | 3 composites/zone (Oct) (6 trawl sites, 4 zones) |
| *Fish Tissues* | offshore | 2 | Hook & Line/Trap | 3 | annual | 1 | 6 | muscle tissue contaminants [g] | 24 | 3 composites/zone (Oct) (2 rig-fishing sites/zones) |
| Totals | | | | | | | 3494 | | 12,124 | |

[a] T, F, E = total coliform, fecal coliform, and enterococcus bacteria (n = 3 parameters) requried at shore and kelp stations

[b] E = enterococcus only required at offshore stations

[c] CTD profile = depth, temperature, salinity, dissolved oxygen, light transmittance (transmissivity), chlorophyll a, pH, density (n = 8 parameters)

[d] CTD profile = depth, temperature, salinity, dissolved oxygen, light transmittance (transmissivity), chlorophyll a, pH, density, and CDOM (n = 9 parameters)

[e] Sediment constituents = sediment grain size, total organic carbon, total nitrogen, sulfides, metals, PCBs, chlorinated pesticides, PAHs, BOD (n = 9 parameter categories; see NPDES permit for complete list of constituents; BOD = voluntary)

[f] Fish tissue contaminants (liver) = lipids, PCBs, chlorinated pesticides, metals (n = 4 parameter categories; see NPDES permit for complete list of constituents)

[g] Fish tissue contaminants (muscle) = lipids, PCBs, chlorinated pesticides, metals (n = 4 parameter categories; see NPDES permit for complete list of constituents)

# Table 4

NPDES-permit mandated receiving waters sampling effort for the South Bay Ocean Outfall Monitoring Program, excluding resamples, QA/QC analyses (e.g., duplicate/split samples), or special studies.

| Monitoring Component | Location | Number of Stations | Sample Type | Discrete No. Samples/Site | Sampling Frequency | Sampling Times/Yr | Discrete No. Samples/Yr | Parameters | No. "Samples" Analyzed/Yr | Notes |
|---|---|---|---|---|---|---|---|---|---|---|
| Water Quality | shore | 11 | Seawater - Bacti | 1 | weekly | 52 | 572 | T, F, E [a] | 1716 | 1 sample/station |
| *Microbiology* & *Oceanographic Conditions* | kelp | 3 | Seawater - Bacti | 3 | 5x/month | 60 | 540 | T, F, E [a] | 1620 | 3 depths/station |
| | | 3 | CTD | 1 | 5x/month | 60 | 180 | CTD profile [b] | 1440 | 1 cast/station (1-m batch avg samples) |
| | offshore | 25 | Seawater - Bacti | 3 | monthly | 12 | 900 | T, F, E [a] | 2700 | 3 depths/station |
| | (n=37) | 37 | CTD | 1 | monthly | 12 | 444 | CTD profile [c] | 3996 | 1 cast/station (1-m batch avg samples) |
| | | 28 | TSS | 3 | monthly | 12 | 1008 | TSS | 1008 | 3 depths/station |
| | | 28 | Oil & Grease | 1 | monthly | 12 | 336 | O&G | 336 | 1 depth/station |
| Sediment Quality | offshore | 27 | Grab | 1 | semiannual | 2 | 54 | sediment constituents [d] | 432 | 1 grab/station (Jan, Jul) |
| Benthic Macrofauna | offshore | 27 | Grab | 2 | semiannual | 2 | 108 | community structure | 108 | 2 replicate grabs/station (Jan, Jul) |
| Demersal Fishes & Invertebrates | offshore | 7 | Trawl | 1 | quarterly | 4 | 28 | community structure | 28 | 1 trawl/station |
| Bioaccumulation *Fish Tissues* | offshore | 7 | Trawl | 3 | semiannual | 2 | 42 | liver tissue contaminants [e] | 210 | 3 composites/station (Apr, Oct) (trawl sites) |
| | offshore | 2 | Hook & Line/Trap | 3 | semiannual | 2 | 12 | muscle tissue contaminants [e] | 60 | 3 composites/station (Apr, Oct) (rig-fishing sites) |
| "Regional Survey" | | | | | | | | | | |
| Sediment Quality | random array | 40 | Grab | 1 | annual | 1 | 40 | sediment constituents [d] | 320 | 1 grab/station (Jul) |
| Benthic Macrofauna | random array | 40 | Grab | 1 | annual | 1 | 40 | community structure | 40 | 2 replicate grabs/station (Jul) |
| Totals | | | | | | | 4304 | | 14,014 | |

[a] T, F, E = total coliform, fecal coliform, and enterococcus bacteria (n = 3 parameters)

[b] CTD profile = depth, temperature, salinity, dissolved oxygen, light transmittance (transmissivity), chlorophyll a, pH, density (n = 8 parameters)

[c] CTD profile = depth, temperature, salinity, dissolved oxygen, light transmittance (transmissivity), chlorophyll a, pH, density, CDOM (n = 9 parameters)

[d] Sediment constituents = sediment grain size, total organic carbon, total nitrogen, sulfides, metals, PCBs, chlorinated pesticides, PAHs (n = 8 parameter categories; see NPDES permit for complete list of constituents)

[e] Fish tissue contaminants = total lipids, metals, PCBs, chlorinated pesticides, PAHs (n = 5 parameter categories; see NPDES permit for complete list of constituents)

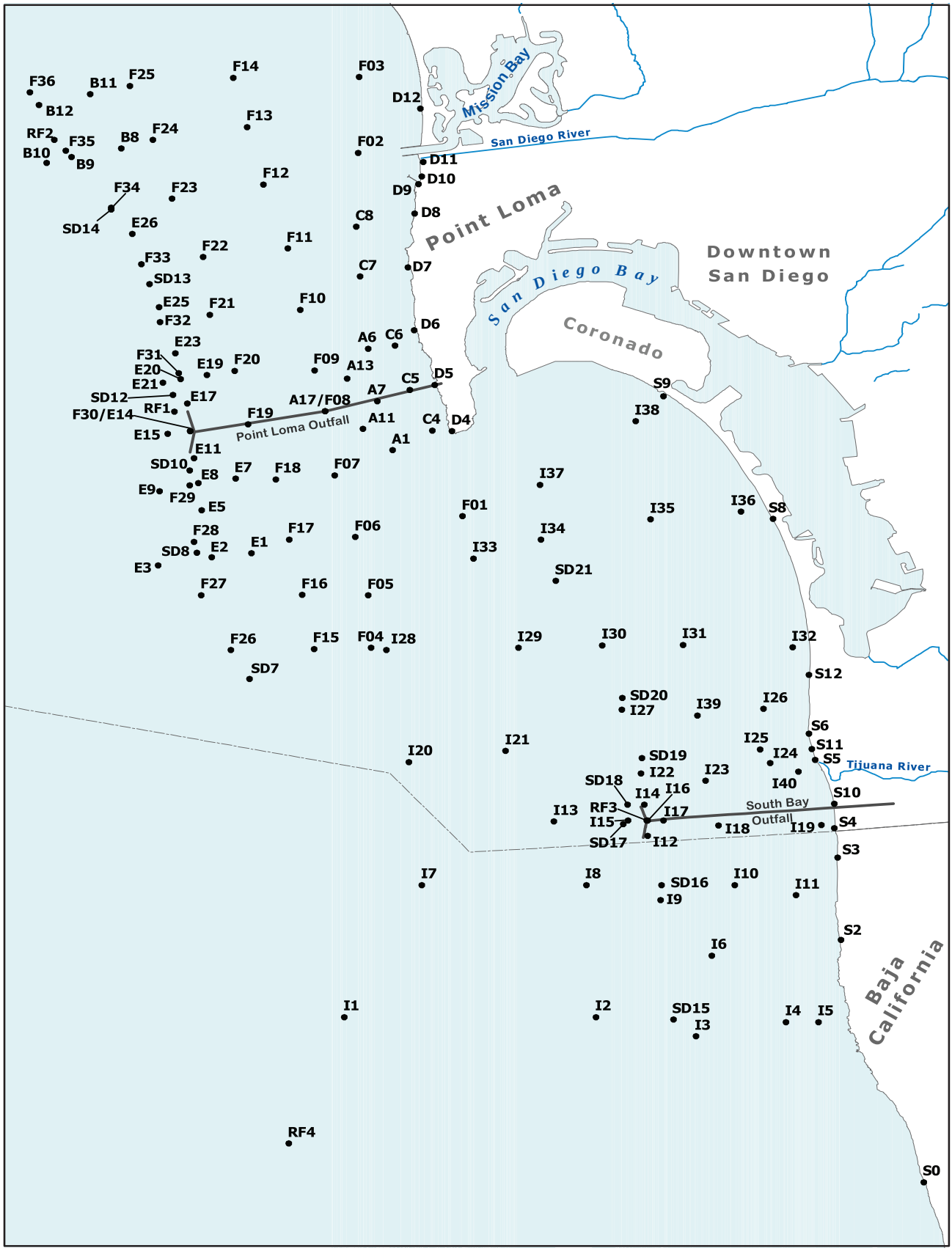



**Figure 1**
Regular fixed monitoring stations for the Point Loma and South Bay Ocean Outfall Monitoring Programs.

## Table 5

Toxicity testing effort for the Point Loma and South Bay Ocean Outfall Monitoring Programs. Listed effort excludes accelerated testing requirements (e.g., triggered by Notice of Violation), additional QA/QC procedures, or special studies.

| Testing Component | Location/ Project | Sample Type | No. samples | Sampling Frequency | Sampling Times/Yr | No. test Species | Effluent/ Ref Tox Tests/Yr | Total Tests/Yr | Endpoints | Dilutions per bioassay | Notes |
|---|---|---|---|---|---|---|---|---|---|---|---|
| **Point Loma** | | | | | | | | | | | |
| Acute toxicty | PLWTP | final effluent | 1 | semi-annual | 2 | 1 | 2 + 2 Ref Tox | 4 | survival | 5 + control | species = mysid |
| | (One-time screening) | final effluent | 1 | 3 x per 2 yrs | 3 x per 2 yrs | 2 | 6+ 6 Ref Tox per 2 yrs | 12 per 2 yrs | survival | 5 + control | screening spp: mysids & topsmelt |
| Chronic toxicity | PLWTP | final effluent | 1 | monthly | 12 | 2 | 24 + 24 Ref Tox | 48 | sensitive lifestage | 5 + control | species = red abalone & giant kelp |
| | (Biennial screening) | final effluent | 1 | 3 x per 2 yrs | 3 x per 2 yrs | 3 | 9+ 9 Ref Tox per 2 yrs | 18 per 2 yrs | sensitive lifestage | 5 + control | screening spp: giant kelp, red abalone, topsmelt |
| **South Bay** | | | | | | | | | | | |
| Acute toxicty | SBWRP | final effluent | 1 | quarterly | 12 | 1 | 12 + 12 Ref Tox | 24 | survival | 5 + control | species = topsmelt |
| | (Biennial screening) | final effluent | 1 | 3 x per 2 yrs | 3 x per 2 yrs | 2 | 6 + 6 Ref Tox per 2 yrs | 12 per 2 yrs | survival | 5 + control | screening spp: mysids & topsmelt |
| | SBWRP/ IWTP | comb. effluent | 1 | quarterly | 4 | 1 | 4 + 4 Ref Tox | 8 | survival | 5 + control | species = topsmelt |
| | (Biennial screening) | comb. effluent | 1 | 3 x per 2 yrs | 3 x per 2 yrs | 2 | 6 + 6 Ref Tox per 2 yrs | 12 per 2 yrs | survival | 5 + control | screening spp: mysids & topsmelt |
| Chronic toxicity | SBWRP | final effluent | 1 | monthly | 12 | 1 | 12 + 12 Ref Tox | 24 | sensitive lifestage | 5 + control | species = red abalone |
| | (Biennial screening) | final effluent | 1 | 3 x per 2 yrs | 3 x per 2 yrs | 3 | 9 + 9 Ref Tox per 2 yrs | 18 per 2 yrs | sensitive lifestage | 5 + control | screening spp: giant kelp, red abalone, topsmelt |
| | SBWRP/ IWTP | comb. effluent | 1 | quarterly | 4 | 1 | 4 + 4 Ref Tox | 8 | sensitive lifestage | 5 + control | species = red abalone |
| | (Biennial screening) | comb. effluent | 1 | 3 x per 2 yrs | 3 x per 2 yrs | 3 | 9 + 9 Ref Tox per 2 yrs | 18 per 2 yrs | sensitive lifestage | 5 + control | screening spp: giant kelp, red abalone, topsmelt |

Comb. Effluent = combined SBWRP + IWTP effluent samples
Ref Tox = Reference Toxicant Test
Sensitive lifestage endpoints: (1) red abalone = development; (2) giant kelp = germination and growth

## Table 6

Number of discrete samples collected and analyzed by the EMTS Division Laboratory for NPDES permit-related activities during 2010. NA = not applicable; WCS = Wastewater Chemistry Services.

| | Number of Samples Collected | | Number of Analyses per Sample Type | |
|---|---|---|---|---|
| **Sample Type** | **Regular** | **QC** | **Regular** | **QA** |
| Sediment Grabs | | | | |
| Particle Size Subsample | 146[a] | NA | (performed by WCS) | |
| Chemistry Subsamples | 628[a,e] | NA | (performed by WCS) | |
| Benthic Infauna Grabs | 244[a] | NA | 244[a] | 30[a] |
| Otter Trawl | 40 | NA | 40 | NA |
| Fish Tissue | 66 | NA | (performed by WCS) | |
| Water Quality | | | | |
| CTD Casts | 1292[b] | NA | 9612[f] | NA |
| Microbiology | 5002[b,d] | 284 | 13,737[c] | 816 (dups)[c]<br>156 (splits)[c] |
| Suspended Solids | 924 | 88 | (performed by WCS) | |
| Oil and Grease | 308 | 66 | (performed by WCS) | |
| $N\text{-}NH_3$ | 143 | NA | 143 | NA |
| Toxicology | | | | |
| Acute Bioassay | 10 | NA | 14 | 13 |
| Chronic Bioassay | 64 | NA | 64 | 40 |

[a] includes Old Outfall special study stations

[b] includes Bight'08 samples

[c] includes three types of analyses (total coliform, fecal coliform and enterococcus)

[d] includes resamples

[e] PLOO stations have five subsamples per grab; all other stations have four

[f] includes up to nine analyses per cast (depth, temperature, salinity, dissolved oxygen, light transmittance, chlorophyll *a*, pH. density, CDOM)

Bight'98, Bight'03 and Bight'08 regional monitoring efforts in 1998, 2003 and 2008, respectively. During these programs, the City's regular sampling and analytical effort may be reallocated as necessary with approval of the RWQCB and U.S. EPA. As with the special studies described above, the regional monitoring efforts are typically subject to QA/QC procedures similar to those for routine monitoring data, although these projects also do not conform to the same analysis and reporting schedules. Thus, the details and results of the current Bight'08 project efforts are not included in this report unless otherwise indicated.

# Summary of Work Performed in 2010

In 2010, a total of 8867 discrete samples and subsamples were collected by EMTS staff, including samples collected as part of permit-mandated special studies (Table 6). Of these, 438 (~5%) represent quality control (QC) samples such as field duplicates. In addition, 1055 quality assurance (QA) tests were conducted to validate the quality of specific analyses such as macrofauna sorting, microbiological analyses and toxicity tests. The results of the QA/QC activities presented in the following sections support the accuracy and precision of the resultant data and validate their use in permit-mandated monitoring or environmental testing and reporting. These include: (1) intercalibration of the

## Table 7

Summary of the CTD intercalibration casts performed during 2010. For each parameter measured by the CTDs, data include mean difference (MeanΔ), maximum difference (MaxΔ), cast number (i.e., 1, 2, or 3), and depth (m) at which the maximum difference occurred.

| Parameter | MeanΔ | MaxΔ | Cast No. | Depth |
|---|---|---|---|---|
| Temperature (°C) | 0.03 | 0.61 | 3 | 10 |
| Salinity (ppt) | 0.01 | 0.11 | 3 | 10 |
| DO (mg/L) | 0.10 | 0.79 | 3 | 13 |
| pH | 0.01 | 0.03 | 1 | 0–110[a] |
| Transmissivity (%) | 1.61 | 3.29 | 2 | 25 |
| Chlorophyll *a* (µg/L) | 0.07 | 0.20 | 3 | 15 |

[a] MaxΔ for pH occurred throughout the entire cast (0–110 m)

Conductivity-Temperature-Depth (CTD) instruments used to sample water quality parameters; (2) results of the bacteriological quality assurance procedures; (3) results of the macrofaunal community sample resorts; (4) results of toxicology quality assurance procedures.

### CTD Intercalibration Exercise

An in-house CTD intercalibration exercise is conducted annually in order to ensure consistency between the two different CTD instruments used to collect the water column profiling data for the City's ocean monitoring program. These two Sea-Bird Electronics Model 25 CTDs were used during the intercalibration exercise, which took place on September 16, 2010. The instrument designated as Unit #3 was a combination CTD/carousel sampler, while Unit #4 was a stand-alone CTD unit. The two CTDs were attached to each other with similar probes aligned during the exercise and deployed to a depth of 100 m three different times. After the three casts were completed, comparisons of six different measured parameters (i.e., temperature, salinity, dissolved oxygen, pH, chlorophyll *a*, transmissivity) and one calculated parameter (density) were performed to assess whether deviations between the instruments and sensors were within acceptable limits (see City of San Diego, in prep).

The results of the 2010 annual intercalibration exercise are summarized in Table 7 and Figure 2, and compared to the results from previous years in Table 8. Comparisons of temperature, salinity, pH, chlorophyll, and dissolved oxygen (DO) values demonstrated acceptable variability between CTDs (i.e., $\leq$0.1 units). However, results from the transmissometer probes displayed slightly more variability between CTD units than in previous years (1.6%). This increase in variability was most likely due to sensor drift of the probes on Unit #4 following its last service.

In addition to the annual in-house intercalibration exercise, each CTD probe is calibrated individually. The temperature, pressure, conductivity and DO probes are calibrated semi-annually by Sea-Bird Electronics at their facility. The fluorometer and transmissometer probes are calibrated annually and CDOM is calibrated semi-annually by Wetlabs at their facility. The transmissometer is also calibrated in-house every six months. The DO probes are calibrated monthly in-house to check for sensor drift. Although the pH probe is factory-calibrated each year, it is also calibrated prior to each monitoring cruise. The pH sensors are serviced in-house when showing slow response times by replacing the electrode component of the sensor. The electrodes are kept in service for a maximum of six months.

## Table 8

Results of annual intercalibration exercises for CTD instruments over the past five years. Values are the differences between Unit #3 and Unit #4 averaged over all depths (0–110 m).

| Parameter | 2010 | 2009 | 2008 | 2007 | 2006 |
|---|---|---|---|---|---|
| Temperature (°C) | 0.03 | 0.07 | 0.05 | 0.03 | 0.06 |
| Salinity (ppt) | 0.01 | 0.02 | 0.01 | 0.01 | 0.01 |
| DO (mg/L) | 0.10 | 0.44 | 0.54 | 0.14 | 0.34 |
| pH | 0.01 | 0.02 | 0.04 | 0.06 | 0.05 |
| Transmissivity (%) | 1.61 | 0.47 | 0.87 | 0.80 | 0.39 |
| Chlorophyll *a* (µg/L) | 0.07 | 0.49 | 0.10 | 0.25 | 0.11 |

### Bacteriological Quality Assurance Analyses

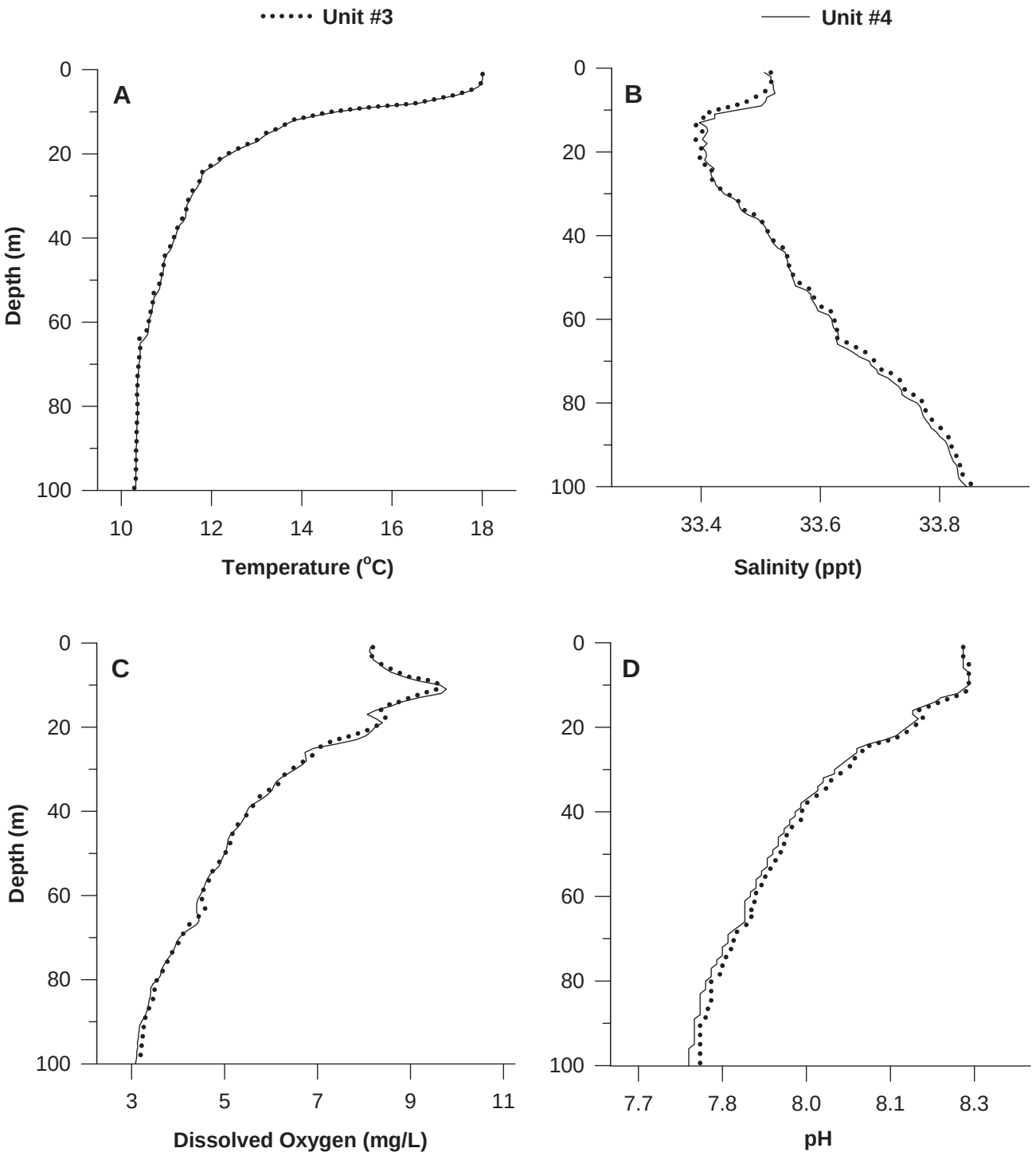
Duplicate and split bacteriological analyses are run throughout the year as quality assurance checks to measure variability between samples and analyst precision, respectively. Duplicate analyses are obtained by taking two separate samples at a given station in the field and then analyzing them in exactly the same way. Split analyses are obtained by taking aliquots of a single field sample and then having two different staff analysts perform the dilutions, filtration, and plating. During 2010, duplicate analyses were performed on approximately 6% of the water quality samples (i.e., $n = 284$ samples and 816 analyses), while split analyses were performed on four to five samples a month (i.e., $n = 52$ samples and 156 analyses). The raw data for these analyses have been reported previously in the monthly receiving waters monitoring reports for each ocean monitoring program.

The sign test (Gilbert 1987) was used to statistically compare the results from the paired duplicate and split analyses performed from January – December 2010 (Table 9). When matched pairs of samples are used, the sign test assumes that the probability of observing samples with differing plate counts is equally distributed among positive (sample A > sample B) and negative (sample A < sample B) results. Samples that do not differ (i.e., $A - B = 0$) are ignored. Results from both the duplicate field sample and split samples were not significantly different ($p > 0.05$) for each of the three tested parameters (i.e., total coliforms, fecal coliforms, enterococcus), indicating low variability between samples, that the measurements performed by each laboratory analyst were repeatable, and that techniques among analysts did not vary significantly.

In addition to these duplicate and split sample analyses, the MML conducts monthly comparisons of bacterial colony counts to quantify the counting precision of each analyst. These comparisons include counts completed by pairs of analysts, where any two analysts must fall within 10% of each other. This calculation is known as the Relative Percent Difference (RPD). In 2010, all of the total and fecal coliform count comparisons were within the required RPD. For enterococcus counts, 2 out of 80 comparisons had an RPD greater than 10%.

### Macrofaunal Community – Resort Analysis

Labatory analysis of benthic macrofaunal (or infaunal) samples involve three processes: sample washing and preservation, sample sorting, and identification and enumeration of all organisms. Quality control of




## Figure 2

Comparison of results from CTD Units #3 and #4 from one representative cast made during the 2010 CTD intercalibration exercise. Data include cast profiles for (A) temperature, (B) salinity, (C) dissolved oxygen, (D) pH, (E) transmissivity, (F) chlorophyll *a*, and (G) density.

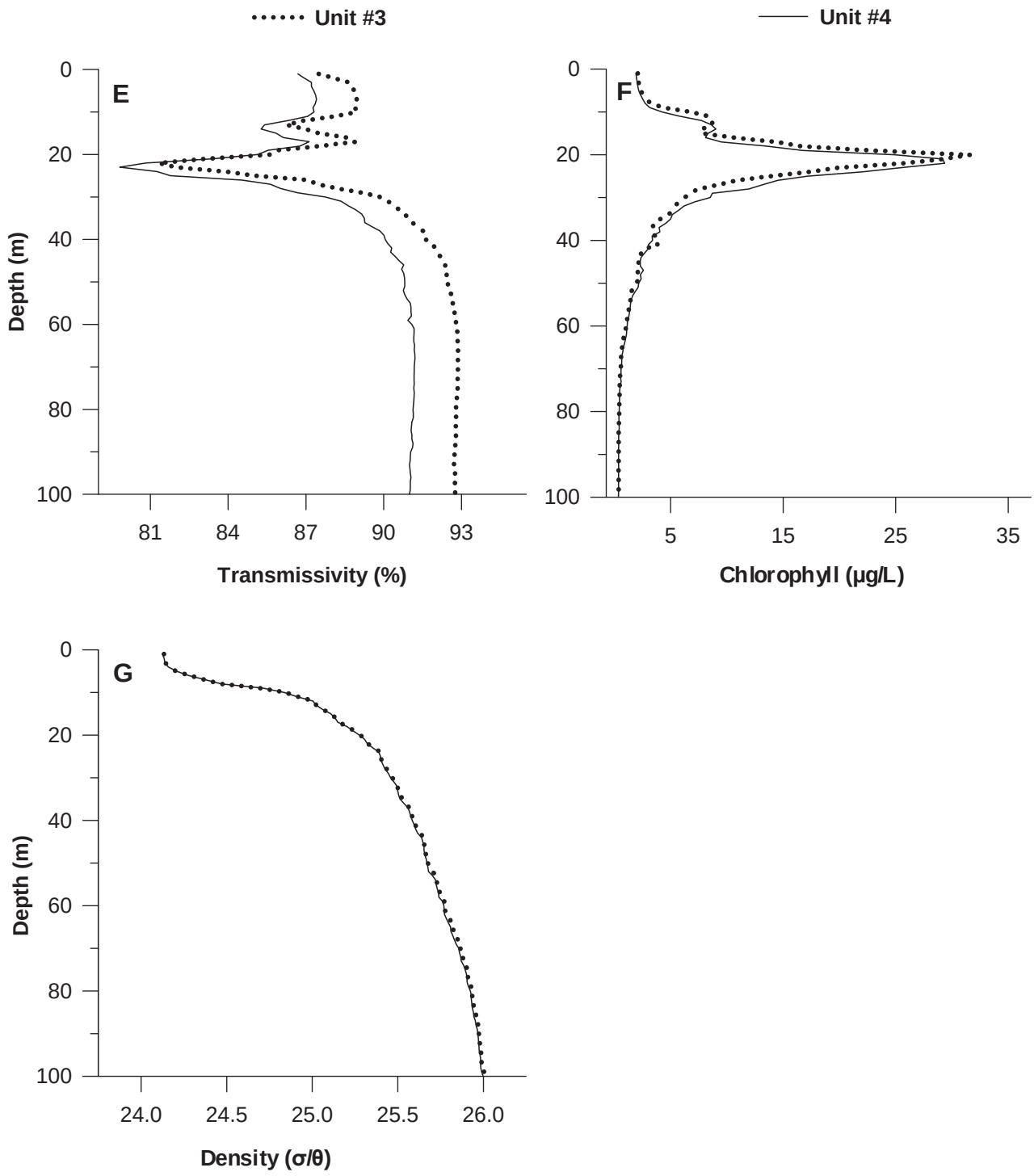



**Figure 2** *continued*

sorting is essential to assure the value of the subsequent steps in the sample analysis process. The sorting of benthic samples is contracted to an outside laboratory, with a 95% removal efficiency expected. Ten percent of the sorted samples from each technician (sorter) are subject to resorting as QA for the contract. The original sorting of a sample fails the QA criteria level if the resorted sample contains more than 5% of the total abundance of organisms from that sample. Failure requires the re-sorting of all samples previously sorted by that sorter. The resort results for the period from January and July 2010 are shown in Table 10. The percentages of animals found in all analyzed samples were ≤5.0% of the total sample abundance.

## Table 9

Summary of bacteriological QA analyses for the Point Loma and South Bay Ocean Outfall Monitoring Programs conducted during 2010. The paired samples were compared using the sign test (see Gilbert 1987) at a $p = 0.05$ level of significance.

| Sample Type | Parameter | N | B | $Z_b$ | $p$ | $H_o$ |
|---|---|---|---|---|---|---|
| Duplicate | Total coliform | 150 | 72 | -0.490 | > 0.05 | Accept |
| | Fecal coliform | 67 | 30 | -0.855 | > 0.05 | Accept |
| | Enterococcus | 60 | 36 | 1.549 | > 0.05 | Accept |
| Split | Total coliform | 25 | 15 | 1.000 | > 0.05 | Accept |
| | Fecal coliform | 21 | 12 | 0.655 | > 0.05 | Accept |
| | Enterococcus | 24 | 11 | -0.408 | > 0.05 | Accept |

$H_o$ = The probability of observing positive and negative differences in plate counts between paired samples is equal (see text)
N = Number of sample pairs with different colony counts; samples without differences are not considered
B = The number of positive differences between pairs
$Z_b$ = Sign test outcome

### Toxicology Quality Assurance Analyses

The Toxicology Laboratory routinely conducts reference toxicant testing as a part of the quality assurance program. A reference toxicant is a standard chemical used to measure the sensitivity of the test organisms in order to establish confidence in the toxicity data obtained from the test material. A specific reference toxicant is used for each test method, and the material is chosen from a list developed by the U.S. EPA. The reference toxicant is purchased from a supplier in aqueous form (stock solution), and the supplier must verify the concentration of the stock solution and provide written documentation of such analysis.

In most instances, a toxicity test with a reference toxicant is performed to assess the sensitivity of the test organisms at the same time the test material (e.g. effluent) is evaluated. A control chart containing no fewer than 20 of the most recent reference toxicant test results for each test method is maintained by the QA officer and is used to monitor test organism sensitivity. Results from a minimum of 19 of the most recent 20 reference toxicant tests must fall within the control chart boundaries (two standard deviations of the mean). Failure to do so triggers an investigation of animal supply, reference toxicant stock quality, and laboratory practices. Additional testing is also conducted to determine whether an exceedance is anomalous or if remedial measures are needed. All NPDES mandated tests conducted with the affected animals are to be flagged, reviewed for anomalous responses, and, in certain cases, repeated with a new batch of animals. In 2010, all reference toxicant control charts met the acceptability criteria.

Also in 2010, the Toxicology Laboratory conducted whole effluent toxicity tests on samples collected at the PLWTP and the SBWRP. A set of concurrent standard reference toxicant tests was conducted with each toxicity test. These tests were mandated by the City's NPDES permits, and the results were included in monthly reports submitted to the RWQCB, U.S. EPA Region IX, State Department of Public Health, and the San Diego County Department of Public Health. Results from these tests will also be summarized and submitted to the agencies above as part of Wastewater Chemistry's annual report for these facilities.

## Table 10

Results of benthic macrofauna sample resort analyses conducted during 2010 for the Point Loma (PLOO and Old Outfall stations) and South Bay (SBOO and Regional stations) Ocean Outfall Monitoring Programs. Percent = (the # of animals found in the resorted sample/the total sample abundance) X 100; [1] and [2] indicate sample replicate number; * indicates samples that have not been completed yet.

| Quarter | Station | Percent | Station | Percent |
|---|---|---|---|---|
| | PLOO | | SBOO | |
| Jan-10 | B-9[1] | 0.00 | I-1[2] | 0.00 |
| | E-7[2] | * | I-10[1] | 0.00 |
| | E-19[1] | * | I-10[2] | 0.00 |
| | E-20[1] | 0.00 | I-18[1] | 0.00 |
| | E-23[2] | 0.00 | I-21[2] | 0.36 |
| | | | I-35[1] | 0.00 |
| Jul-10 | B-8[1] | * | I-2[2] | 0.79 |
| | E-5[1] | * | I-8[1] | 0.00 |
| | E-9[2] | 0.00 | I-9[1] | 0.00 |
| | E-11[2] | 0.00 | I-13[2] | 0.00 |
| | E-17[1] | 0.00 | I-21[1] | 0.00 |
| | E-26[2] | 0.00 | I-33[2] | 0.00 |
| | | | I-35[1] | 0.91 |
| | Old Outfall | | Regional 2010 | |
| Jul-10 | A-9 | * | 8006 | 0.00 |
| | | | 8015 | 0.00 |
| | | | 8025 | 0.00 |
| | | | 8028 | 0.00 |
| | | | 8037 | 0.00 |

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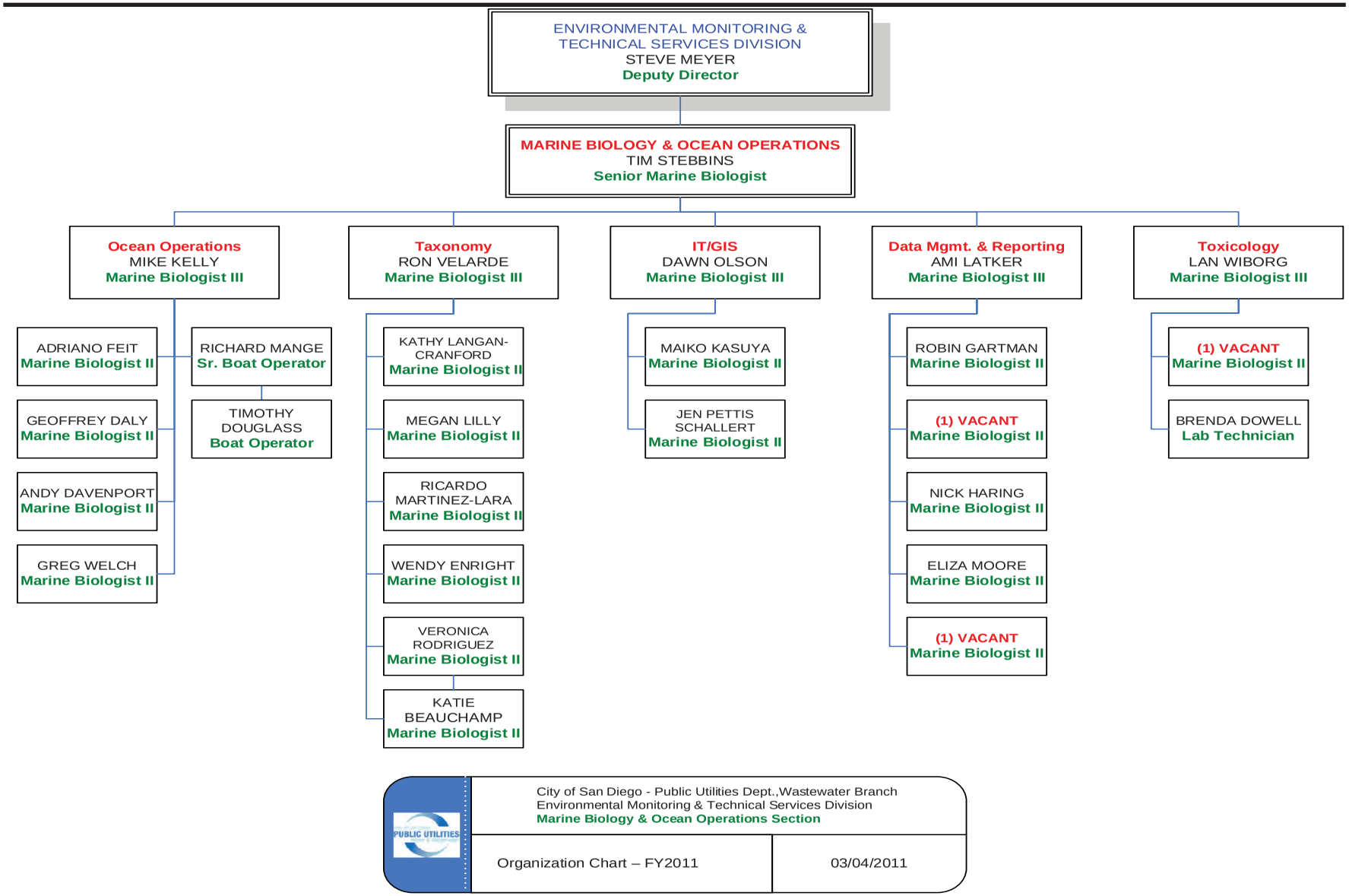
# APPENDIX A

## Organizational Charts

# Appendix A.1

Organizational chart for the Marine Biology and Ocean Operations section of the Environmental Monitoring and Technical Services Division Laboratory.



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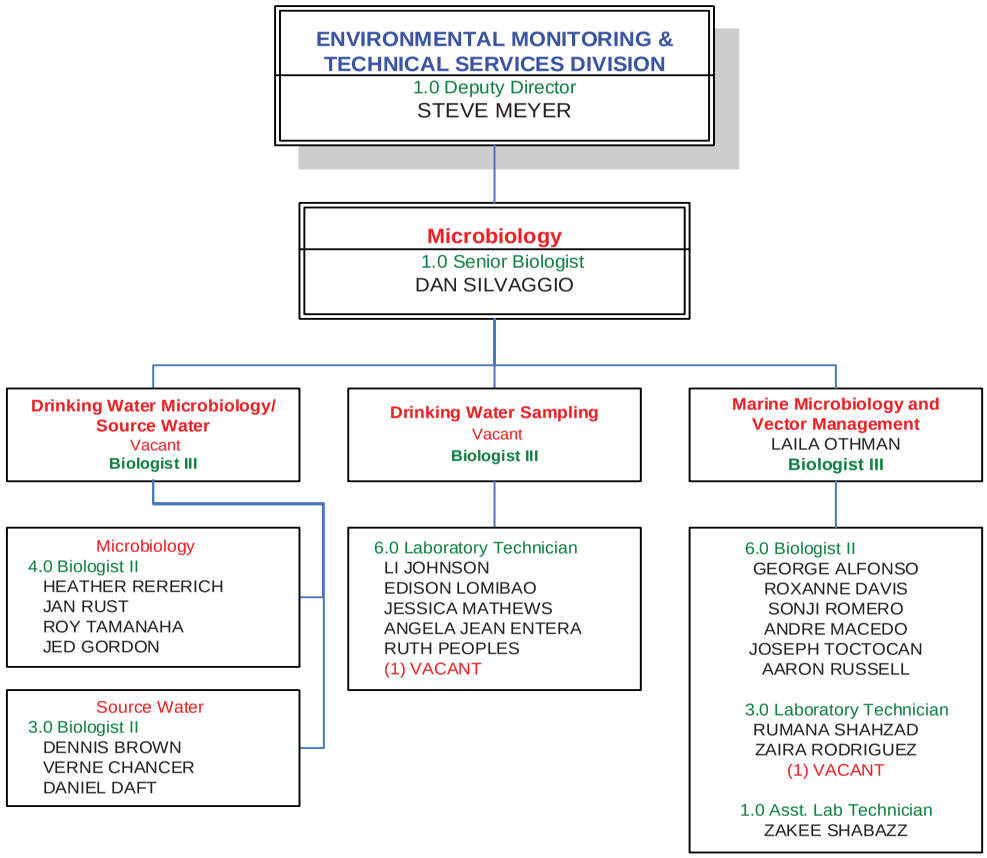
# Appendix A.2

Organizational chart for the Microbiology section of the Environmental Monitoring and Technical Services Division Laboratory.




City of San Diego-Public Utilities Dept., Wastewater Branch
Environmental Monitoring & Technical Services Division
**Marine Microbiology & Vector Management**

| Organization Chart – FY2011 | 02/22/2011 |
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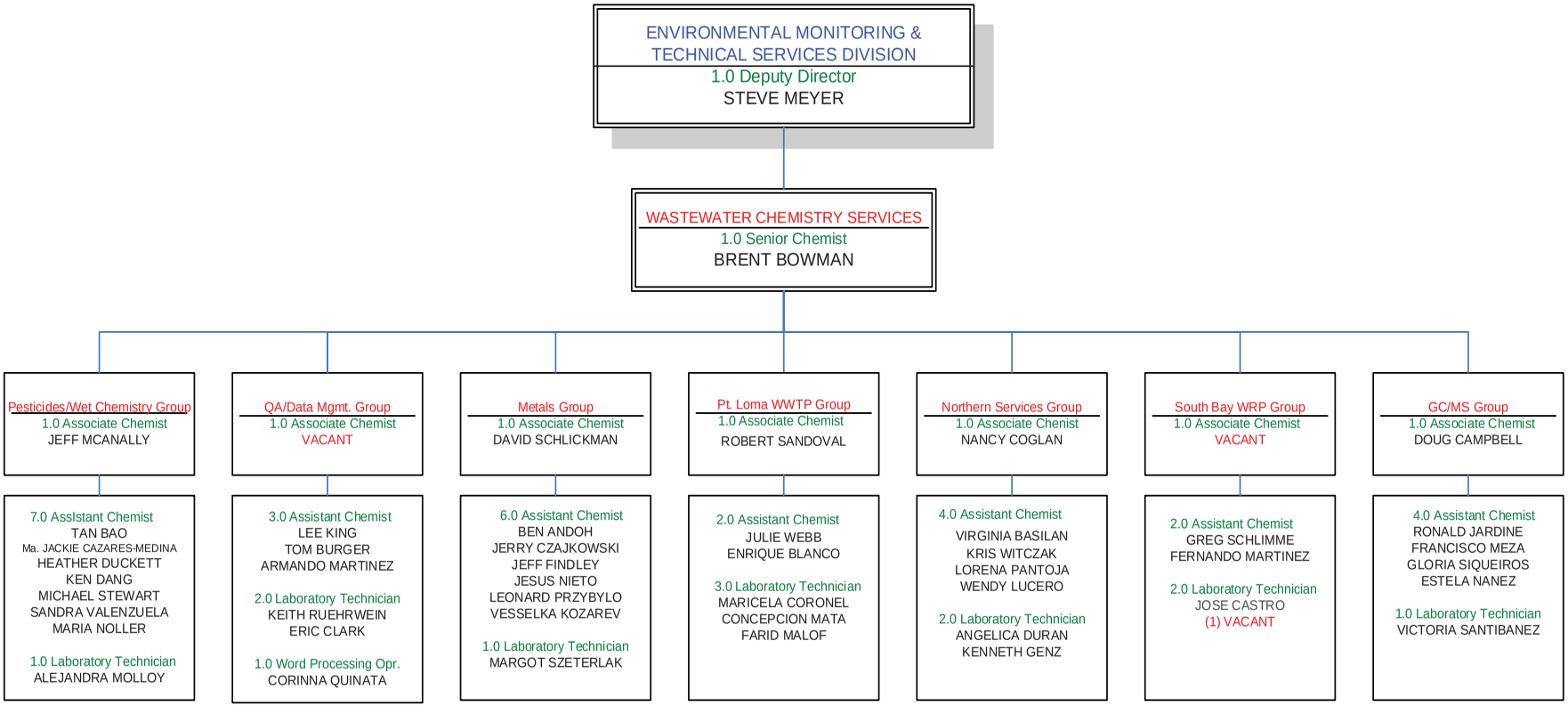
# Appendix A.3

Organizational chart for the Wastewater Chemistry Services section of the Environmental Monitoring and Technical Services Division Laboratory.




City of San Diego – Public Utilities Dept. Wastewater Branch
Environmental Monitoring & Technical Services Division
**Wastewater Chemistry Services**

| Organization Chart – FY2011 | 02/22/2011 |
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